Total synthesis of a keramamide

Jennifer A. Sowinski and Peter L. Toogood*

Willard H. Dow Laboratory, Department of Chemistry, University of Michigan, Ann Arbor, MI 48109-1055, USA. E-mail: peter.toogood@wl.com

Received (in Corvallis, OR, USA) 3rd March 1999, Accepted 15th April 1999

The first total synthesis of a molecule possessing the structure proposed for keramamide J is described, providing data indicating that the structure of this natural product should be re-examined.

Keramamide J (KJ; 1)¹ is the simplest member of a class of *Theonella* natural products that collectively exhibit cytotoxic,¹⁻³ anti-fungal⁴ and anti-oxidant activity.⁵ To study the biological activities of these molecules, we required a general synthetic route that would be amenable to the synthesis of all members of this series.⁴ Herein, we report our progress towards this goal, including the first total synthesis of a keramamide possessing structure 1.



Our synthetic plan for constructing KJ is shown in Scheme 1.⁶ A convergent route was devised, employing four fragments of comparable complexity (**3**–**6**). We decided to mask the electrophilic keto-amide moiety⁷ as a protected α -hydroxy carbonyl group until the conclusion of the synthesis to avoid nucleophilic attack at this center. In addition, we elected to attempt ring closure between the amine of fragment **3** and the C-terminus of the alanine residue, which was anticipated to be less prone to epimerization than the tryptophan residue and easier to couple than the α -hydroxy acid. Late incorporation of side chain fragment **6** was selected to increase convergency and minimize potential side reactions during the critical macrocyclization reaction.

Thiazole 7^8 was extended *via* reduction of the ester to an aldehyde and Wittig olefination to introduce the *E*-double bond (Scheme 2). Saponification of the ethyl ester, followed by coupling to L-tryptophan allyl ester provided fragment **4**. To obtain the hexanoic acid fragment **3**, enone **9** (Scheme 3) was



Scheme 1



Scheme 2 *Reagents and conditions*: i, NaOH, MeOH; ii, BOP, Et₃N, HN(Me)OMe; iii, LiAlH₄; iv, Ph₃P=CHCO₂Et (70% for 4 steps); v, NaOH, MeOH; vi, L-Trp-OAllyl, Ph₂POCl (68% for 2 steps).



Scheme 3 Reagents and conditions: i, NaBH₄, CeCl₃; ii, separate (76% for 2 steps); iii, SEMCl, 2,6-lutidine; iv, O₃, DMS; v, NaClO₂, isobutene; vi, H₂, Pd/C; vii, FmocOSu, aq. NaHCO₃ (66% for 5 steps).

prepared from L-isoleucine via displacement of the corresponding Weinreb amide with vinyl magnesium bromide. Reduction of this enone with NaBH₄ under the Luche conditions⁹ provided a 4:1 mixture of alcohols which could be separated by column chromatography. Protection of the major product $(10)^{10}$ as its (trimethylsilyl)ethoxymethyl (SEM) acetal, followed by a two step oxidation of the double bond, provided the carboxylic acid. Hydrogenolytic cleavage of the Cbz group and subsequent treatment with FmocOSu[†] gave compound 3. Deprotection of fragment 4, followed by condensation with acid 3, gave the corresponding tripeptide in 77% yield (Scheme 4). Subsequent cleavage of the allyl ester and coupling to dipeptide 5 gave linear precursor to the KJ macrocycle 11. Following deprotection of the C-terminus, and removal of the Fmoc group under standard conditions, macrocyclization proceeded smoothly to provide cyclic peptide 12 in 34-51% overall yields from intermediate 11.

To prepare the KJ side chain, (*S*)-glycidol was oxidized to glycidic acid using $RuCl_3$ and $NaIO_4$,¹¹ then coupled with L-isoleucine methyl ester to give peptide **13** as a single diastereomer (Scheme 5). Attack of the oxirane by azide ion at



Scheme 4 Reagents and conditions: i, HCl, Et₂O; ii, **3**, DCC, HOBt, $Pr^{i}_{2}EtN$; iii, Pd(PPh₃)₄, dimedone; iv, **5**, DCC, HOBt, $Pr^{i}_{2}EtN$ (56% for 4 steps); v, Pd(PPh₃)₄, dimedone; vi, Et₂NH; vii, (PhO)₂PON₃, NaHCO₃ (51% over 3 steps).



Scheme 5 Reagents and conditions: i, RuCl₃, NaIO₄; ii, L-Ile-OMe, DCC, HOBt (36% for 2 steps); iii, NaN3, MgSO4; iv, LiOH; v, CsCO3, BnBr; vi, TESCl, imidazole (48% for 4 steps); vii, PPh₃, H₂O; viii, p-O₂NC₆H₄-OCHO; ix, H₂, Pd/C (79% for 3 steps).

the less substituted position, followed by transesterification and protection of the hydroxy function produced azido peptide 14. Staudinger reduction of the azide to a primary amine,¹² followed by formylation with *p*-nitrophenyl formate and hydrogenolytic cleavage of the ester, provided fragment 6.

Treatment of macrocycle 12 with HCl in Et₂O-MeOH-CHCl3 deprotected the amine and secondary alcohol, and fragment $\hat{\mathbf{6}}$ was attached using HATU^{\dagger} to produce alcohol 15 (Scheme 6).¹³ Oxidation of the alcohol was performed under mild, non-acid conditions, using IBX⁺ in DMSO.¹⁴ Finally, removal of the TES group was achieved by stirring the peptide over Amberlite IR-120 suspended in EtOAc. Purification of the final product by silica gel chromatography followed by reversed-phase HPLC provided material that exhibits ¹H, ¹³C, ¹H-¹H COSY and inverse detected ¹H-¹³C HMQC NMR spectra consistent with the proposed structure 1, and a high resolution mass spectrum corresponding to the expected molecular formula.



Scheme 6 Reagents and conditions: i, HCl, MeOH; ii, 6, HATU, 2,4,6-collidine (48% for 2 steps); iii, IBX, DMSO; iv, Amberlite IR-120 (95% for 2 steps).

From the NMR data, it is apparent that our synthetic keramamide is not the same compound reported by Kobayashi and co-workers.^{1,15} Comparison of the NMR spectra leads us to conclude that this synthetic keramamide and KJ are configurational isomers. In support of the assignment of structure 1 to the synthetic product, a very close correlation is observed between the spectral data for this compound and the data published for KF (2; Table 1), which differs only in the replacement of the tryptophan in structure 1 by Z-didehydrotryptophan. In particular, the ¹H and ¹³C chemical shifts at C-9 and C-13 for these two compounds are in excellent agreement (¹H $\Delta \delta \leq 0.06$, ¹³C $\Delta \delta \leq 1.3$ ppm). In contrast, the published data for KJ more closely resemble the data for keramamide G (KG) which is epimeric to KF at carbon-13.^{1,16} We note that the degradation conditions used by Kobayashi to determine the absolute configuration at carbon-13 in KJ have been found previously to cause epimerization at this center in a closely related molecule and possibly could have been misleading.³ Degradation of the synthetic keramamide under milder conditions using 30% H₂O₂

Table 1 Selected ¹H and ¹³C NMR resonances, and optical rotations published for keramamides F, G, J and observed for compound 1

	$\delta_{ m H}$		$\delta_{ m C}$		
	H-9	H-13	C-9	C-13	$[\alpha]^{25\ a}_{\mathrm{D}}$
Keramamide G	4.81	5.49	53.7	56.7	+10.0
Keramamide J	4.75	5.49	53.7	56.1	+8.4
Compound 1	5.31	5.19	51.4	61.0	-10.0
Keramamide F	5.33	5.25	51.7	59.7	-25

degrees cm

and 0.1 M NaOH (room temperature, overnight), followed by 6 M HCl (110 °C, 24 h), produced L-isoleucine containing less than 10% D-allo-isoleucine by chiral HPLC analysis, supporting assignment of the L-configuration to carbon-13 in the synthetic material.¹⁷ Upon standing in aqueous solution, the synthetic material partially converted ($\sim 10\%$) to a new product possessing ¹H NMR resonances that match those observed for KJ, consistent with the notion that these two molecules differ at a single, epimerizable center.

Based on the preceding analysis, we conclude that our synthesis proceeded as intended to correctly provide a molecule possessing structure 1. This work strongly indicates that the structure of KJ should be revised. However, since no natural KJ is presently available,15 structural re-assignment will require either re-isolation or total synthesis of the correct structure.

This work was supported by NSF grant CHE-93221233, and by the donors to the Petroleum Research Fund, administered by the American Chemical Society.

Notes and references

† Abbreviations: HATU = O-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate, IBX = 1-hydroxy-1,3-dihydro-1,2-benziodoxol-3-one 1-oxide, BOP = benzotriazol-1-yloxytris(dimethylamino)phosphonium hexafluorophosphate, FmocOSu = N-(fluoren-9-ylmethoxycarbonyloxy)succinimide, HOBt = 1-hydroxybenzotriazole hydrate.

- 1 J. Kobayashi, F. Itagaki, H. Shigemori, T. Takao and Y. Shimonishi, Tetrahedron, 1995, 51, 2525.
- 2 F. Itagaki, H. Shigemori, M. Ishibashi, T. Nakamura, T. Sasaki, and J. Kobayashi, J. Am. Chem. Soc., 1992, 57, 5540.
- 3 N. Fusetani, T. Sugawara, S. Matsunaga and H. Hirota, J. Am. Chem. Soc. 1991, 113, 7811.
- S. P. Gunasekara, S. A. Pomponi and P. J. McCarthy, J. Nat. Prod., 1994. 57. 79.
- 5 J. Kobayashi, F. Itagaki, H. Shigemori, M. Ishibashi, K. Takahashi, M. Ogura, S. Nagasawa, T. Nakamura, H. Hirota, T. Ohta and S. Nozoe, J. Am. Chem. Soc., 1991, 113, 7812.
- 6 Preliminary work on this synthesis has been published: J. A. Sowinski and P. L. Toogood, Tetrahedron Lett., 1995, 36, 67.
- 7 Examples of other ketoamide containing peptides see: N. Fusetani, S. Matsunaga, H. Matsumoto and Y. Y. Takebayashi, J. Am. Chem. Soc., 1990, 112, 7053; M. Hagihara and S. L. Schreiber, J. Am. Chem. Soc., 1992, 114, 6570; S. Toda, C. Kotake, T. Tsuno, Y. Narita, T. Yamasaki and M. Konishi, J. Antibiot., 1992, 45, 1580; T. Aoyagi, M. Nagai, K. Ogawa, F. Kojima, M. Okada, T. Ikeda, M. Hamada and T. Takeuchi, J. Antibiot., 1991, 44, 949; M. Nagai, K. Ogawa, Y. Muraoka, H. Naganawa, T. Aoyagi and T. Takeuchi, J. Antibiot., 1991, 44, 956.
- 8 J. A. Sowinski and P. L. Toogood, J. Org. Chem., 1996, 61, 7671.
- 9 J.-L. Luche, J. Am. Chem. Soc., 1978, 100, 2226; J.-L. Luche, L. Rodriguez-Hahn and P. J. Crabbé, J. Chem. Soc., Chem. Commun., 1978. 601.
- 10 The major product from this reduction was identified through its conversion to the corresponding oxazolidinone (NaH, DMF) and ¹H NMR spectral comparison with related literature compounds, See for example, T. Ibuka, H. Habashita, A. Otaka, N. Fujii, Y. Oguchi, T. Uyegara and Y. Yamamoto, J. Org. Chem., 1991, 56, 4370.
- 11 C. H. Behrens and K. B. Sharpless, J. Org. Chem., 1985, 50, 5696.
- 12 H. Staudinger and J. Meyer, Helv. Chim. Acta, 1919, 2, 635; N. Knouzi, M. Voultier and R. Carrie, Bull. Soc. Chim. Fr., 1985, 815.
- 13 L. A. Carpino, J. Am. Chem. Soc., 1993, 115, 4397; L. A. Carpino and A. El-Faham, J. Org. Chem., 1994, 59, 695; L. A. Carpino, A. El-Faham, C. A. Minor and F. Albericio, J. Chem. Soc., Chem. Commun., 1994, 201; L. A. Carpino, A. El-Faham and F. Albericio, Tetrahedron Lett., 1994, 35, 2279; L. A. Carpino and A. El-Faham, J. Org. Chem., 1995. 60. 3561.
- 14 M. Frigerio, M. Santagostino, S. Sputore and G. Palmisano, J. Org. Chem., 1995, 60, 7272.
- 15 Authentic samples of KJ and KF are not available. We thank Professor Kobayashi for providing a copy of the ¹H NMR spectrum for natural KI
- 16 Chemical degradation of KG converts the homo-Ile fragment to Disoleucine indicating that KG possesses the (R) configuration at both C-13 and C-15.
- A peak corresponding to L-alanine was also detected in this analysis, 17 indicating that this residue did not epimerize during the cyclization reaction.

Communication 9/01928F